

Research paper**Anticholinesterase, Antioxidant and Nitric Oxide Scavenging Activity of the Aqueous Extract of Some Medicinal Plants**

Aims: Enhancement of cholinergic activity and reduction of oxidative stress by scavenging free radicals such as nitric oxide are well recognized therapeutic approaches in several pathological conditions. We evaluated the anticholinesterase, antioxidant and nitric oxide scavenging activity of the aqueous extracts of *Ocimum basilicum*, *Curcuma longa* and *Solanum nigrum*.

Study design: Experimental

Place and duration of study: Delhi Institute of Pharmaceutical Sciences & Research, Delhi University, New Delhi, India between January 2008 and December 2008.

Methodology: The aqueous extracts of the rhizome of *Curcuma longa*, berries of *Solanum nigrum* and seeds of *Ocimum basilicum* were authenticated by HPTLC fingerprinting. The anticholinesterase activity of these extracts was estimated spectrophotometrically as described by Ellman in 1961 and IC₅₀ was calculated. Total antioxidant capacity of extracts was also estimated spectrophotometrically based on the reduction of molybdenum (Mo) (VI) to Mo(V) by the sample and the subsequent formation of a green phosphate/Mo(V) complex at acidic pH. Ascorbic acid was used as standard. Estimation of nitric oxide scavenging activity of extracts was based on the diazotization reaction.

Results: The anticholinesterase activity (IC₅₀) was observed at the concentrations of 2.73 ± 0.09 , 3.38 ± 0.05 and 3.88 ± 0.11 gram/l for *Solanum nigrum*, *Curcuma longa*, and *Ocimum basilicum* respectively. At these concentrations, maximum antioxidant capacity equivalent to 4.36 ± 0.14 mmol of ascorbic acid was shown by *Curcuma longa*, followed by *Solanum nigrum*, and *Ocimum basilicum*. *Curcuma longa* showed the maximum nitric oxide scavenging activity equivalent to 29.78 ± 1.28 mmol of sodium nitrite followed by *Solanum nigrum* and *Ocimum basilicum*.

Conclusion: Plant derived pharmacological agents may provide an attractive therapeutic option in future for several pathological conditions especially the neurodegenerative diseases due to their anticholinesterase, antioxidant and nitric oxide scavenging properties.

Keywords: Anticholinesterase, Antioxidant, Nitric oxide scavenging activity, Aqueous plant extracts

1. INTRODUCTION

Neurodegenerative disorders such as Alzheimer's and Parkinson's disease are characterized by reduced cholinergic activity in brain [1]. The enzyme cholinesterase which exists as acetylcholinesterase (AChE) and butyrylcholinesterase (BChE) causes hydrolysis of acetylcholine and its inhibition, therefore, plays a key role in enhancing cholinergic activity. Besides reduced cholinergic activity, oxidative stress has also been recognized as a key factor in the pathogenesis of neurodegenerative disorders like Alzheimer's [2]. Oxidation is an integral process in the cellular metabolism and during oxidation, free radicals are produced that have unpaired electron [3-6]. Oxygen and nitrogen atoms with free unpaired

21 electron are highly reactive and produce cellular damage by causing membrane lipid
22 peroxidation and damage to enzymes and DNA [7].

23 Accordingly, the current therapeutic options in neurodegenerative disorders primarily involve
24 the drugs that can increase the cholinergic activity as well as are antioxidants. A variety of
25 anticholinesterases and antioxidants have been used, however, search for more effective
26 and safer agents continues.

27 Several agents from plant origin have been investigated previously for their AchE inhibitory
28 and antioxidant properties. In the present study we evaluated the AchE inhibitory, total
29 antioxidant and NO scavenging activity of the aqueous extracts of the seeds of *Ocimum*
30 *basilicum* (OB), rhizomes of *Curcuma longa* (CL) and berries of *Solanum nigrum* (SN).

31 OB is an annual and perennial herb and shrub that belongs to the family Lamiaceae. It is also
32 known as common basil or the sweet basil. In addition to the essential oils, it is rich in
33 flavonoids and anthocyanins. The different types of OB extracts have been shown to
34 possess antioxidant properties [8]. The essential oils from OB have also been shown to
35 possess AchE inhibitory activity [9]. CL, a perennial herb, is a member of the Zingiberaceae
36 (ginger) family. Curcuminoids from CL have been shown to possess memory enhancing
37 activities in vitro and in vivo models [10]. Curcumin from CL has also been shown to
38 possess nitric oxide (NO) scavenging and antioxidant properties [11, 12]. SN belongs to the
39 family Solanaceae and consists of glycoalkaloids, glycoproteins and saponins. Its green
40 berries and leaves contain glycoalkaloids like solanine and solasodine. The principal
41 alkaloid, solanine, has anticholinesterase action that is attributed to its aglycone solanidine
42 and the fruit extract has been shown to possess antioxidant properties [13]. As stated,
43 various components from various parts of these 3 plants have been evaluated for AchE
44 inhibitory, total antioxidant and NO scavenging properties. In the current study, for the first
45 time, we evaluated these activities of the aqueous extracts of OB seeds, CL rhizome and SN
46 berries.

47

48 **2. MATERIAL AND METHODS**

49

50 **2.1 Plant Extracts**

51 Dried aqueous plant extracts from seeds of OB, rhizome of CL and berries of SN were
52 provided by Promed Exports Private Ltd, India, and all extracts were authenticated by
53 HPTLC finger printing.

54 **2.2 Evaluation of AchE Inhibitory Activity**

55 The AChE inhibitory activity of three extracts was measured according to the method
56 developed by Ellman et al., in 1961 [14]. All estimations were done in triplicates. The
57 concentrations of the tested extracts that inhibited the hydrolysis of substrate by 50% (IC₅₀)
58 was estimated for all three extracts and their combination by a linear regression analysis
59 between the percentage inhibition versus the extract concentrations by using the Microsoft
60 Excel program.

61

62 **2.2.1 Principle of Reaction**

63 Acetylcholine iodide is used as the substrate. When acted upon by the enzyme AChE it
64 breaks down to thiocholine and acetate. Thiocholine is allowed to react with
65 dithiobisnitrobenzoate (DTNB) and this reaction results in the development of a yellow

66 colour. The change in the intensity of yellow color over time, which can be estimated using a
67 UV spectrophotometer, is a measure of the activity of AChE.

68

69 **2.2.2 Preparation of Enzyme and Solutions**

70 Plasma from the venous blood of human volunteers was used as a source of enzyme AChE.
71 Acetylcholine iodide 0.1mM and DTNB 0.3mM solution was prepared in phosphate buffer
72 with pH 7.4. Physostigmine 1mM was prepared in distilled water and was used as reference
73 standard. The aqueous plant extracts were dissolved in distilled water so as to get the
74 desired concentration.

75

76 **2.2.3 Assay**

77 Fifty microliters of plasma was added to the assay tubes containing 3 ml of phosphate buffer
78 and tubes were then incubated for 5 min at 37°C. After incubation, 50 µl of extract sample or
79 reference standard was added. 50 µl of distilled water was added instead of sample for
80 blank. Solutions were again incubated with intermittent shaking. DTNB solution, 100 µl, was
81 now added to the tubes. This was followed by quick addition of 100 µl of acetylcholine
82 iodide. The intensity of color change was measured with spectrophotometer at 412 nm at
83 kinetic mode. Readings were taken at an interval of 15 sec for a total of 3 min.

84

85 **2.2.4 Calculation of enzyme activity**

86 The rate of color change per min was calculated for each reading. The rates were then
87 averaged within each three min run. The rate of reaction was calculated according to
88 following formula:

89

$$\text{Activity (mol/min/l)} = \frac{\text{Change in absorbance} \times \text{Assay volume} \times 1000}{\text{Absorption coefficient} \times \text{Light path} \times \text{Sample volume}}$$

90

91

92 Assay volume = 3.3 ml; Absorption coefficient = 1.36×10^{-4} /M/cm; Sample volume = 0.05
93 ml; Light path = 1cm

94

95 As no enzyme inhibition is taking place in blank the enzyme activity of blank was taken as
96 100%. By comparing with blank, percent enzyme activity and percent inhibition of enzyme
97 activity was calculated.

98

99 **2.3 Evaluation of antioxidant activity**

100 The evaluation of total antioxidant capacity was based on the method described by Prieto *et*
101 *al.*, in 1999 [15]. Each extract was used in the concentration equivalent to IC50 for AchE
102 inhibition. All estimations were done in triplicates.

103

104 **2.3.1 Principle of Reaction**

105 This phosphomolybdenum method is now commonly used in extensive screenings of
106 samples of very different origins and composition in search for powerful antioxidants. The
107 assay is based on the reduction of Mo(VI) to Mo(V) by the sample and the subsequent
108 formation of a green phosphate/Mo(V) complex at acidic pH. The method was optimized and
109 characterized with respect to linearity interval, repetitivity, reproducibility, and molar
110 absorption coefficients for the quantitation of several antioxidants by Prieto *et al.*, in 1999
111 [15].

112

113 **2.3.2 Assay**

114 An aliquot of 0.1 ml of sample solution containing the aqueous extracts, in the same
115 concentrations as for AchE inhibitory activity, was combined with 1 ml of reagent solution
116 containing 0.6M sulfuric acid, 28mM sodium phosphate, and 4mM ammonium molybdate.
117 The solution was incubated in a water bath at 95°C for 90 min. After the samples had cooled
118 to room temperature, the absorbance of the aqueous solution of each was measured at 695
119 nm against a blank. A typical blank solution contained 1ml of reagent solution and the equal
120 volume of water as used for the sample. Incubation was done under the same conditions as
121 the rest of the samples. Ascorbic acid, a water-soluble antioxidant, was used as standard
122 and calibration curve was obtained using various concentrations of ascorbic acid. The
123 antioxidant capacity was expressed as the equivalent of mmols of ascorbic acid.

124

125 **2.4 Evaluation of NO scavenging activity**

126 Evaluation of NO scavenging activity was based on the method described by Griess in 1879
127 [16]. All estimations were done in triplicates.

128

129 **2.4.1 Principle of Reaction**

130 NO in oxygen-containing aqueous solution has a short half-life due to its rapid oxidation. It
131 has been reported that NO in aqueous solution containing oxygen is oxidized primarily to
132 nitrite (NO_2^-) with little or no formation of nitrate (NO_3^-) [17]. So, the NO formation is
133 assessed by measuring NO_2^- . The assay relies on a diazotization reaction. The reaction
134 utilizes sulfanilamide and N-1-napthylethylenediamine dihydrochloride (NED) under acidic
135 conditions. Sodium nitroprusside in aqueous solution at physiological pH spontaneously
136 generates NO [18, 11], which interacts with oxygen to produce nitrite ions that can be
137 estimated by use of Griess reagent. Scavengers of NO compete with oxygen leading to
138 reduced production of nitrite ions [19].

139

140 **2.4.2 Assay**

141 Sodium nitroprusside (5 mM in PBS at pH 7.4) 100 μl solution was mixed with 750 μl of
142 different concentrations of sodium nitrite (10 – 70 mM in water) or the equal volume of
143 extracts and incubated at 25°C for 150 min. After incubation 200 μl of Griess reagent,
144 containing 1% ($w v^{-1}$) sulphanilamide, 0.1% ($w v^{-1}$) NED and 2.5% ($v v^{-1}$) phosphoric acid,
145 was added and the absorbance of the coloured compound formed due to diazotization of
146 nitrite with sulfanilamide and subsequent coupling with NED was read at 546 nm. The linear
147 standard curve was obtained by plotting the mean absorbance for each standard
148 concentration against the sodium nitrite concentration. The standard curve was used to
149 calculate the sodium nitrite (mM) equivalent activity in the test sample.

150

151

152 **3. RESULTS AND DISCUSSION**

153

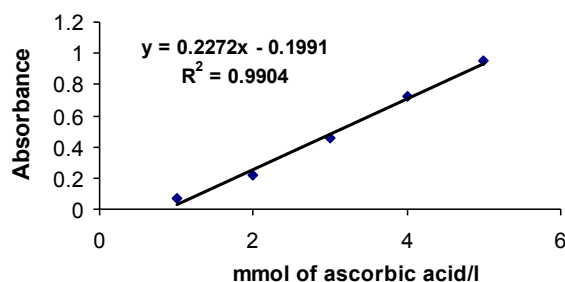
154 **3.1 AchE Inhibitory Activity**

155 The method was first validated and absorbance was measured in the absence of inhibitor as
156 well as in the presence of different concentrations of physostigmine (0.5 – 1.5 mmol). The
157 linearity of method was established. In the presence of physostigmine, a potent
158 anticholinesterase inhibitor, significant inhibition of AchE was observed. Physostigmine in a
159 concentration of 1mM resulted in a 95.25% inhibition of AChE activity. Among the extracts
160 50% of the AchE inhibitory activity (IC_{50}) was observed at the concentrations of 2.73 ± 0.09 ,
161 3.38 ± 0.05 and 3.88 ± 0.11 gm/l for SN, CL and OB respectively. A combination of all three
162 extracts at above concentrations showed 72.25% AchE inhibition.

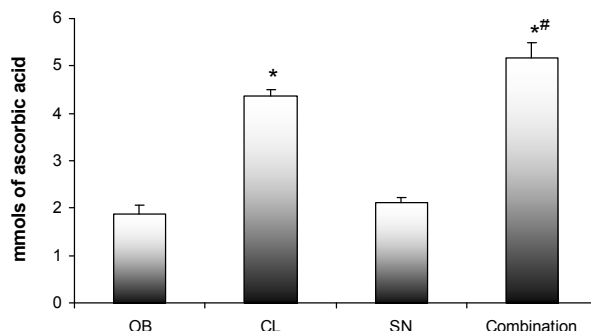
163

164 **3.2 Total Antioxidant Capacity**

165 The phosphomolybdenum assay was performed using ascorbic acid in the concentration
 166 range of 1 to 5 mmol (Fig 1). The antioxidant capacity of the three aqueous extracts was
 167 estimated and expressed as equivalents of mmol of ascorbic acid. Among the three extracts
 168 the maximum antioxidant capacity was shown by CL, which was equivalent to 4.36 ± 0.14
 169 mmol of ascorbic acid followed by SN and OB with a mean value of 2.12 ± 0.11 and $1.88 \pm$
 170 0.18 mmol of ascorbic acid respectively. The antioxidant capacity of CL was significantly
 171 higher as compared to other extracts ($P < 0.001$). The antioxidant capacity of combination
 172 consisting of OB, CL and SN was equivalent to 5.95 ± 0.32 mmol of ascorbic acid. The
 173 antioxidant capacity of herbal combination was significantly higher than all its constituent
 174 extracts ($P < 0.05$ vs CL and $P < 0.001$ vs OB and SN) (Fig 2).
 175
 176
 177



178 Fig 1: Standard curve for ascorbic acid in phosphomolybdenum assay.
 179
 180
 181

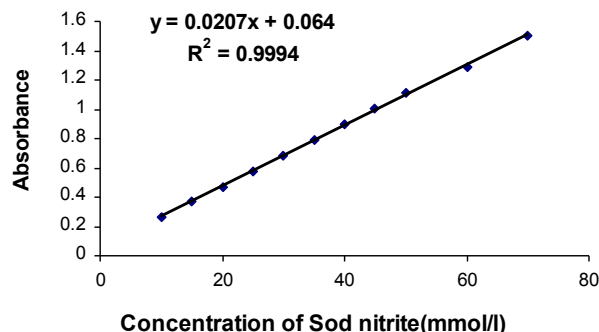


182 Fig 2: Total antioxidant capacity of three extracts and their combination.
 183
 184 * $P < 0.001$ versus OB and SN; # $P < 0.05$ versus CL
 185

186 **3.3 NO Scavenging Activity**

187 The NO scavenging activity of three extracts and their combinations was estimated in a
 188 diazotization reaction. The calibration curve for sodium nitrite (10-70 mmol/l) was used to
 189 calculate the NO scavenging activity of test drugs which was expressed as equivalent to
 190 mmol of sodium nitrite (Fig 3). Among the three aqueous extracts, the CL showed the
 191 maximum NO scavenging activity, which was equivalent to 29.78 ± 1.28 mmol of sodium
 192 nitrite. The NO scavenging activity of SN and OB was equivalent to 11.71 ± 1.84 and 11.34
 193 ± 2.30 mmol of sodium nitrite respectively. The NO scavenging activity of CL was
 194 significantly higher than two other extracts ($P < 0.01$). The NO scavenging activity of the

195 combination of three extracts was equivalent to 39.83 ± 1.82 mmol of sodium nitrite and this
 196 was significantly higher than their constituent extracts ($P < 0.001$ vs OB & SN, $P < 0.05$ vs CL).
 197 (Fig 4)
 198

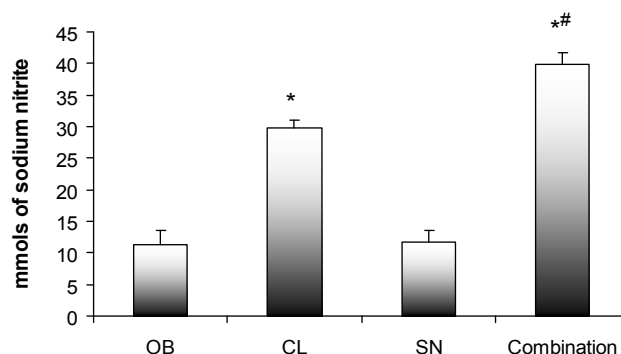


199

200 Fig 3: Diazotization reaction using different concentrations of sodium nitrite

201

202



203 Fig 4: Nitric oxide scavenging activity of three plant extracts and their combination

* $P < 0.001$ versus OB and SN; # $P < 0.05$ versus CL

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

Present study demonstrated AchE inhibitory, antioxidant and nitric oxide scavenging activity of the aqueous extracts of three medicinal plants. Enhancement of cholinergic activity by prolonging the availability of acetylcholine in synaptic clefts is a well recognized therapeutic approach in several pathological conditions especially the neurodegenerative diseases. Inhibition of AchE and butrylcholinesterase (BchE) provides the basis of such therapeutic options. Inhibition of AchE has been shown to enhance cholinergic transmission in brain and additionally it has been observed that AchE inhibition reduces aggregation of β -amyloid and formation of neurotoxic fibrils in Alzheimer's disease [20]. Inhibition of BchE in cases with BchE polymorphism having reduced BchE activity has also been shown to slow down the progression of Alzheimer's disease [21]. Thus, AchE and BchE inhibitors have been recognized as remarkable alternatives [22]. As the Ellman reaction measures both AchE and BchE activity, the extracts evaluated in this study were found to have significant achE and BchE inhibitory activity. Oxidative stress as an underlying pathophysiological process is also well recognized in these neurodegenerative disease processes. ROS are responsible for the damage of cellular bio-molecules such as proteins, enzymes, nucleic acids, lipids and

223 carbohydrates and may adversely affect immune functions [23]. Antioxidants and nitric oxide
224 scavengers, therefore, play a key role by preventing the cellular damage either by
225 scavenging them or by reducing their production. Existing anticholinesterase drugs such as
226 tacrine, donepezil, galantamine and heptylphosphostigmine cause several adverse effects such
227 as hepatotoxicity. Additionally these drugs have short duration of action, low bioavailability,
228 peripheral cholinergic adverse effects and a narrow therapeutic window. Therefore,
229 investigations for newer drugs that possess both AchE inhibitory and antioxidant properties
230 and are safe is extremely important.

231 Historically, active components from plants have provided important sources of new drugs.
232 Since, neurodegenerative diseases such as Alzheimer's have become a public health
233 burden and the currently available drugs have undesirable side-effects, new treatment
234 options based on medicinal plants may be useful therapeutic options.

235

236

237 **4. CONCLUSION**

238

239 The aqueous extracts of the *Curcuma longa* rhizome, *Solanum nigrum* berries and *Ocimum*
240 *basilicum* seeds showed significant anticholinesterase, antioxidant and nitric oxide
241 scavenging properties. New treatment options based on these plant extracts may provide an
242 attractive therapeutic option in future.

243

244

245 **ACKNOWLEDGEMENTS**

246

247 The authors acknowledge the financial support by Department of Science and Technology,
248 Government of India, for carrying out this work.

249

250

251 **COMPETING INTERESTS**

252

253 All authors declare that no competing interests exist.

254

255 **AUTHORS' CONTRIBUTIONS**

256

257 Renu Agarwal, designed the study, performed the experiment and statistical analysis, wrote
258 the protocol, and wrote the first draft of the manuscript.

259 SK Gupta designed the study and protocol and participated in manuscript revision and final
260 approval.

261 Puneet Agarwal participated in writing protocol and manuscript

262 Sushma Srivastava participated in study design, conducting the experiment and manuscript
263 preparation.

264 Renad Alyautdin participated in manuscript revision and final approval.

265 All authors approved the final version of manuscript.

266

267 **REFERENCES**

268 1. Nunes-Tavares N, Santos LE, Stutz B, Brito-Moreira J, Klein WL, Ferreira ST, de
269 Mello FG. Inhibition of choline acetyltransferase as a mechanism for cholinergic
270 dysfunction induced by amyloid- β peptide oligomers. *J Biol Chem.*
271 2012;287(23):19377-85.

272 2. Lee SH, Kim KR, Ryu SY, Son S, Hong HS, Mook-Jung I, Lee SH, Ho WK. Impaired
273 short-term plasticity in mossy fiber synapses caused by mitochondrial dysfunction of

- 274 dentate granule cells is the earliest synaptic deficit in a mouse model of Alzheimer's
275 disease. *J Neurosci.* 2012;33(17):5953-63.
- 276 3. Finkel T, Holbrook NJ. Oxidants, oxidative stress and the biology of ageing. *Nature.*
277 2000;408: 239-47.
- 278 4. Halliwell B. The antioxidant paradox. *Lancet.* 2000;355:1179-80.
- 279 5. Pietta P. Flavonoids as antioxidant. *J Nat Prod.* 2000;63:1035-42.
- 280 6. Visioli F, Keaney Jr JF, Halliwell B. Antioxidants and cardiovascular disease;
281 pancreas or tonics for tired sheep. *Cardiovasc Res.* 2000;47:409-18.
- 282 7. Husain SR, Cillard J, Cillard P. Hydroxyl radical scavenging activity of flavonoids.
283 *Phytochemistry.* 1987;26:2489-97.
- 284 8. Kaurinovic B, Popovic M, Vlaisavljevic S, Trivic S. Antioxidant capacity of *Ocimum*
285 *basilicum* L. and *Origanum vulgare* L. extracts. *Molecules.* 2011;16(9):7401-14.
- 286 9. Orhan, Kartal M, Kan Y, Sener B. Activity of essential oils and individual
287 components against acetyl- and butyrylcholinesterase. *Z Naturforsch C.* 2008;63(7-
288 8):547-53.
- 289 10. Ahmed T, Gilani A H. Inhibitory effect of curcuminoids on acetylcholinesterase
290 activity and attenuation of scopolamine-induced amnesia may explain medicinal use
291 of turmeric in Alzheimer's disease. *Pharmacol Biochem Behav.* 2009;91(4):554-9.
- 292 11. Sreejayan, Rao M N. Nitric oxide scavenging by curcuminoids. *J Pharm Pharmacol.*
293 1997;49:105-7.
- 294 12. Huang HC, Xu K, Jiang Z F. Curcumin-Mediated Neuroprotection Against Amyloid-
295 β -Induced Mitochondrial Dysfunction Involves the Inhibition of GSK-3 β . *J Alzheimers*
296 *Dis.* 2012;32(4):981-96.
- 297 13. Perez RM, Perez JA, Garcia LMD, Sossa HM. Neuropharmacological activity of
298 *Solanum nigrum* fruit. *J. Ethnopharmacol.* 1998;62(1):43-8.
- 299 14. Ellman GL, Callaway E. Erythrocyte cholinesterase-levels in mental patients. *Nature.*
300 1961;192:1216.
- 301 15. Prieto P, Pineda M, Aguilar M. Spectrophotometric quantitation
302 of antioxidant capacity through the formation of a phosphomolybdenum complex:
303 specific application to the determination of vitamin E. *Anal Biochem.* 1999;269(2):
304 337-41.
- 305 16. Griess P. Bemerkungen zu der abhandlung der H.H.Weselsky und Benedikt "Ueber
306 einige azoverbindungen". *Chem Ber.* 1879;12:426-8.
- 307 17. Ignarro LJ, Fukuto JM, Griscavage JM, Rogers NE, Byrns RE. Oxidation of Nitric
308 Oxide in Aqueous Solution to Nitrite but not Nitrate: Comparison with Enzymatically
309 Formed Nitric Oxide From L-Arginine. *Proc Nat Acad Sci.* 199;90:8103-7.
- 310 18. Marcocci L, Maquire JJ, Droy-Lefaix MT, Packer L. The nitric oxide-scavenging
311 properties of *Gingko biloba* extract EGb 761. *Biochem Biophys Res Commun.*
312 1994;201(2):748-55.
- 313 19. Marcocci L, Maquire JJ, Droy-Lefaix MT, Packer L. Antioxidant actions of *Gingko*
314 *biloba* extract EGb 761. *Methods Enzymol.* 1996;234:462-75.
- 315 20. J. R. Hodges, "Alzheimer's centennial legacy: origins, landmarks and the current
316 status of knowledge concerning cognitive aspects," *Brain*, vol. 129, pp. 2811-2822,
317 Nov. 2006.
- 318 21. Loizzo M R, Menichini F, Conforti F, Tundis R, Bonesi M, Saab A M, Statti G A,
319 Cindio B, Houghton PJ, Menichini F, Frega N G. Chemical analysis, antioxidant,
320 anti-inflammatory and anticholinesterase activities of *Origanum ehrenbergii* Boiss
321 and *Origanum syriacum* L. essential oils. *Food Chem.* 2009;117:174-80.
- 322 22. Orhan, Sener B, Choudhary MI, Khalid A. Acetylcholinesterase and
323 butyrylcholinesterase inhibitory activity of some Turkish medicinal plants. *J*
324 *Ethnopharmacol.* 2004;91:57-60.

- 325 23. Nilsson J, Stegmark R, Åkesson B. Total antioxidant capacity in different pea (*Pisum*
326 *sativum*) varieties after blanching and freezing. Food Chemistry. 2004;86(4):501–7.
327
328
329